

SAT-23-P-BFT(R)-07

ANTIMICROBIAL ACTIVITY OF *BACILLUS METHYLOTROPHICUS* 47
AGAINST PHYTOPATHOGENIC FUNGI

Rositsa Denkova

Department of Biochemistry and molecular biology
University of Food Technologies, Plovdiv, BG
E-mail: rositsa_denkova@mail.bg

Bogdan Goranov

Desislava Teneva

Zapryana Denkova

Hristo Marinov

Department of Microbiology
University of Food Technologies, Plovdiv, BG
E-mail: dr.eng.bgoranov@gmail.com;
E-mail: desi_gerinska@yahoo.com;
E-mail: zdenkova@abv.bg;
E-mail: hr_marinov@abv.bg

Georgi Kostov

Department of Wine and Brewing
University of Food Technologies, Plovdiv, BG
E-mail: george_kostov2@abv.bg

Antimicrobial activity of *Bacillus methylotrophicus* 47 against phytopathogenic fungi: A Gram-positive, spore-forming, rod-shaped bacteria strain was isolated from healing spring waters in the village of Mineralni Bani, Haskovo region. It was identified as *Bacillus methylotrophicus* by the application of modern molecular-genetic methods – ARDRA-analysis and sequencing of 16S rRNA gene. The strain was designated as *Bacillus methylotrophicus* 47. The strain was cultured under aerobic or anaerobic conditions at 30°C or 37°C at different pH values of the medium (LBG-broth or MPB). Its antimicrobial activity on the growth of phytopathogenic fungi of *Aspergillus* sp. was examined. *Bacillus methylotrophicus* 47 showed high inhibitory activity against fungi of *Aspergillus* sp. after culturing under aerobic or anaerobic conditions at pH values in the range from pH = 5.0 to pH = 8.0. It was determined that the inhibitory activity was due to the synthesis of an antibiotic substance of a peptide nature with Rf value of 0.65.

Key words: *Bacillus methylotrophicus*, antifungal, phytopathogenic, antimicrobial, *Aspergillus*.

INTRODUCTION

Representatives of the genus *Bacillus* produce substances with antimicrobial activity of different chemical nature. The antimicrobial agents include peptides, amino sugars, phospholipids, etc. [4].

According to Leifert et al., 1995; Yazgan et al., 2001; Montesinos, 2007 [1, 2, 5] the antimicrobial compounds produced by strains of the genus *Bacillus* exhibit antifungal activity against phytopathogenic fungi.

In recent years aflatoxins in food and the protection of the population from their carcinogenic effect are the subject of growing social interest. The development of preparations with high antifungal activity against *Aspergillus flavus* would allow to process both soil and plant individuals. This in turn would enable the production of safe food.

The purpose of the present work was to study the antifungal activity of a *Bacillus methylotrophicus* strain against pathogenic fungi of the genus *Aspergillus*.

MATERIALS AND METHODS

Microorganisms. *Bacillus methylotrophicus* 47 isolated from spring waters in the village of Yabalkovo, Haskovo region, Bulgaria and identified to species level by sequencing the 16S rDNA

was used in the present study. As test microorganisms were used the following plant isolates: *Aspergillus niger*, *Aspergillus awamori*, *Aspergillus flavus*.

Media

a. MPB. Composition (g/dm³): peptone - 10; NaCl - 5; meat extract - 3; glucose - 10. Sterilization - 20 min at 121 °C.

b. LBG-broth. Composition (g/dm³): tryptone - 10; yeast extract - 5; NaCl - 10; glucose - 10. Sterilization - 20 min at 121 °C.

c. LBG-agar. Composition (g/dm³): tryptone - 10; yeast extract - 5; NaCl - 10; glucose - 10 agar - 15. Sterilization - 20 min at 121 °C.

d. MPA. Composition (g/dm³): tryptone - 10; yeast extract - 5; NaCl - 10; glucose - 10. Sterilization - 20 min at 121 °C.

Methods of analysis

Culturing of *Bacillus methylotrophicus* 47

The culturing of *Bacillus methylotrophicus* 47 was carried out in 500 cm³ Erlenmeyer flasks that contained 10 cm³ of culture medium (LBG-broth or MPB) under aerobic conditions obtained by a rotary shaker (220 min⁻¹) and under anaerobic conditions (static culturing in a thermostat) at a temperature of 37 °C for 24-48h. The inoculum was 1% (v/v) 18-hour vegetative inoculum. The culturing was conducted at various pH values of the broth media (LBG-broth or MPB) ranging from pH=4.5 to pH=8.0.

Preparation of a sterile filtrate of the culture suspension

After the culturing the culture suspension was centrifuged at 3500 min⁻¹ for 15 min to separate the biomass. The supernatant was filtered through a membrane filter (pore size 0,45 µm). The biomass was washed twice with saline solution and was adjusted to the initial sample volume.

Determination of the antimicrobial activity – well-diffusion method

To determine the antimicrobial activity of *Bacillus methylotrophicus* 47 against phytopathogenic fungi were prepared three variants of each sample: culture suspension (CS), biomass in saline solution (BSS) and acellular supernatant (ASN), obtained after 24-48 hours of culturing of *Bacillus methylotrophicus* 47. The antimicrobial activity was investigated against *Aspergillus niger*, *Aspergillus awamori*, *Aspergillus flavus*. For the preparation of the fungal spore suspensions the respective test fungal microorganism was grown in an incubator at 30 °C on LBG-agar for 3 to 7 days. Spore suspensions of each of the test microorganisms (10⁶-10⁷ cfu/cm³) were prepared and they were used to inoculate Petri dishes (pour plating) with LBG-agar, the final spore concentration in the inoculated LBG-agar medium was 10⁵cfu/cm³. After the solidification of medium wells (d=6mm) were prepared. 0,06 cm³ of the CS, BSS and ASN were pipetted in the wells and the Petri dishes were incubated at 30 °C or 37 °C for 24h to 48 h. The experiments were performed in quadruplicate. The results as average inhibition zones in mm were reported on the 24th or 48th hour of incubation at 30 °C or 37 °C.

Thin layer chromatography (TLC)

TLC for the separation of the produced antibiotic substances in the culture suspension (CS), biomass in saline solution (BSS) and the acellular supernatant (ASN) was performed. The system used was butanol: acetic acid: water (3: 1: 1 v/v/v). Chromatographic silica gel plates COF₂₅₄ 20x20 cm (Merck, Germany) were used. The plates were placed in a chromatography chamber in a layer with height of 0,5 cm. At the start of the chromatographic plate with a micropipette were placed 50µl of the culture suspension (CS), the acellular supernatant (ASN) or the biomass in saline solution (BSS). The chromatograms were run in duplicate. After the thin layer chromatography, the chromatograms were dried at 60 °C for about 15min and were sprayed with ninhydrin solution (to prove the presence of peptide antibiotics according to the method of Niederwieser, 1972 [3]). After

the spraying of the chromatograms pink spots appeared. The Rf - values of the active substances were determined.

RESULTS AND DISCUSSION

The most appropriate temperature range for the growth of *Bacillus methylotrophicus* 47 is in the temperature range of 30-37°C. Thus, its antimicrobial action was determined at two temperatures - 30 °C and 37 °C, after static or dynamic culturing in two broths – LBG-broth and MPB. The culturing was conducted at various pH values of the broth media ranging from pH=4.5 to pH=8.0. The inhibitory activity was determined by the well-diffusion method. The results as inhibition zones in mm were reported on the 24th or 48th hour of incubation at 30 °C or 37 °C.

The test results for the antimicrobial activity of *Bacillus methylotrophicus* 47 cultured under anaerobic conditions at 30 °C are presented in Table 1.

Table 1.

Antimicrobial activity of *Bacillus methylotrophicus* 47 cultured under anaerobic conditions for 48 hours against phytopathogenic fungi at incubation temperature of 30 °C

Media		Inhibition zones, mm		
		Test-microorganism		
		<i>Aspergillus niger</i> , 3×10^5 cfu/cm ³	<i>Aspergillus awamori</i> , $1,2 \times 10^5$ cfu/cm ³	<i>Aspergillus flavus</i> , $1,12 \times 10^5$ cfu/cm ³
LBG pH = 4,5	ASN	17	25	27
	CS	18	25	35
	BSS	20	25	34
LBG pH = 5,0	ASN	30	25	28
	CS	37	35	37
	BSS	37	37	35
LBG pH = 6,0	ASN	30	30	28
	CS	39	37	32
	BSS	39	39	28
LBG pH = 7,0	ASN	32	30	35
	CS	36	35	40
	BSS	27	35	33
LBG pH = 8,0	ASN	28	25	30
	CS	37	27	37
	BSS	34	32	27
MPB pH = 4,5	ASN	20	25	23
	CS	28	32	27
	BSS	27	28	25
MPB pH = 5,0	ASN	27	30	23
	CS	33	32	28
	BSS	33	30	27
MPB pH = 6,0	ASN	27	20	22
	CS	30	24	23
	BSS	27	27	24
MPB pH = 7,0	ASN	30	32	30
	CS	34	37	35
	BSS	34	34	30
MPB pH = 8,0	ASN	30	30	27
	CS	30	35	35
	BSS	27	32	40

B. methylotrophicus 47 exhibited strong antifungal activity, the largest inhibition zones being reported at samples obtained by culturing at pH=7.0 in LBG-broth and MPB under anaerobic conditions for 48 h (Table 1). The inhibition zones were very clear which is a proof of the fungicidal action of *B. methylotrophicus* 47.

The samples obtained after the aerobic culturing of the strain demonstrate large inhibition zones (Table 2). High antifungal activity against *Asp. niger* and *Asp. awamori* was reported in the samples obtained after culturing in LBG-broth and MPB with pH = 5.0. The high antimicrobial activity of the acellular supernatant (ASN) and the biomass in saline solution (BSS) indicated that the antimicrobial components were associated with the cell surfaces as well as secreted into the medium.

Table 2.

Antimicrobial activity of *Bacillus methylotrophicus* 47 cultured under aerobic conditions for 48 hours against phytopathogenic fungi at incubation temperature of 30 °C

Media		Inhibition zones, mm		
		Test-microorganism		
		<i>Aspergillus niger</i> , $3 \times 10^5 \text{ cfu/cm}^3$	<i>Aspergillus awamori</i> , $1,2 \times 10^5 \text{ cfu/cm}^3$	<i>Aspergillus flavus</i> , $1,12 \times 10^5 \text{ cfu/cm}^3$
LBG pH = 4,5	ASN	20	30	35
	CS	26	32	34
	BSS	18	23	37
LBG pH = 5,0	ASN	32	33	37
	CS	43	42	37
	BSS	40	42	40
LBG pH = 6,0	ASN	27	35	30
	CS	36	40	37
	BSS	31	40	32
LBG pH = 7,0	ASN	24	37	35
	CS	32	43	43
	BSS	34	35	37
LBG pH = 8,0	ASN	28	23	42
	CS	32	32	42
	BSS	32	34	40
MPB pH = 4,5	ASN	29	39	26
	CS	30	37	30
	BSS	33	35	34
MPB pH = 5,0	ASN	28	30	35
	CS	37	37	37
	BSS	30	33	43
MPB pH = 6,0	ASN	25	22	25
	CS	31	27	32
	BSS	30	27	31
MPB pH = 7,0	ASN	33	30	34
	CS	36	35	34
	BSS	36	32	34
MPB pH = 8,0	ASN	35	32	23
	CS	30	35	32
	BSS	37	33	35

When the temperature of culturing was raised to 37°C the inhibitory activity of the strain was also amended. From the samples obtained after the 48-hour anaerobic culturing of *B. methylotrophicus* 47 the highest inhibitory activity was established in the samples with culturing in LBG-broth medium with pH=7.0 and in MPB with pH=6.0 (Table 3).

Table 3.

Antimicrobial activity of *Bacillus methylotrophicus* 47 cultured under anaerobic conditions for 48 hours against phytopathogenic fungi at incubation temperature of 37 °C

Media		Inhibition zones, mm		
		Test-microorganism		
		<i>Aspergillus niger</i> , $3 \times 10^5 \text{ cfu/cm}^3$	<i>Aspergillus awamori</i> , $1.2 \times 10^5 \text{ cfu/cm}^3$	<i>Aspergillus flavus</i> , $1.12 \times 10^5 \text{ cfu/cm}^3$
LBG pH = 4,5	ASN	12	22	17
	CS	12	23	17
	BSS	13	20	20
LBG pH = 5,0	ASN	25	24	15
	CS	27	27	17
	BSS	34	27	20
LBG pH = 6,0	ASN	25	30	34
	CS	30	32	36
	BSS	27	36	39
LBG pH = 7,0	ASN	30	27	33
	CS	25	32	33
	BSS	34	34	35
LBG pH = 8,0	ASN	36	30	27
	CS	30	27	25
	BSS	35	32	32
MPB pH = 4,5	ASN	26	30	29
	CS	26	28	30
	BSS	32	30	30
MPB pH = 5,0	ASN	28	25	28
	CS	34	35	33
	BSS	29	32	33
MPB pH = 6,0	ASN	30	32	30
	CS	30	32	32
	BSS	36	35	36
MPB pH = 7,0	ASN	30	26	22
	CS	28	30	22
	BSS	45	25	23
MPB pH = 8,0	ASN	30	27	25
	CS	30	32	30
	BSS	33	27	30

In the aerobic culturing of *B. methylotrophicus* 47 at a temperature of 37 °C high antimicrobial activity was determined in LBG-broth with pH=6.0 and in MPB with pH=7.0, the diameter of the inhibition zone exceeding 40 mm (Table 4) in the culture suspension (CS) and the biomass in saline solution (BSS), which reaffirmed the fact that the antimicrobial compounds were associated with the microbial cells. This showed that *B. methylotrophicus* 47 could grow under anaerobic conditions and in both media at pH=6-7 the samples demonstrated high antifungal activity. Higher antimicrobial activity was determined in the samples obtained after the aerobic cultivation at 37 °C in both media at pH=6.0 (Table 4).

Table 4.
 Antimicrobial activity of *Bacillus methylotrophicus* 47 cultured under aerobic conditions for 24 hours against phytopathogenic fungi at incubation temperature of 37 °C

Media		Inhibition zones, mm		
		Test-microorganism		
		<i>Aspergillus niger</i> , $3 \times 10^5 \text{ cfu/cm}^3$	<i>Aspergillus awamori</i> , $1,2 \times 10^5 \text{ cfu/cm}^3$	<i>Aspergillus flavus</i> , $1,12 \times 10^5 \text{ cfu/cm}^3$
LBG pH = 4,5	ASN	8	22	16
	CS	10	17	15
	BSS	11	20	13
LBG pH = 5,0	ASN	30	35	35
	CS	22	25	28
	BSS	27	27	30
LBG pH = 6,0	ASN	35	25	25
	CS	30	37	32
	BSS	25	32	35
LBG pH = 7,0	ASN	32	24	25
	CS	27	30	32
	BSS	30	32	30
LBG pH = 8,0	ASN	34	22	23
	CS	34	32	25
	BSS	25	29	25
MPB pH = 4,5	ASN	32	22	24
	CS	34	28	30
	BSS	32	25	30
MPB pH = 5,0	ASN	32	30	32
	CS	32	40	40
	BSS	32	37	37
MPB pH = 6,0	ASN	28	32	25
	CS	37	44	37
	BSS	37	34	35
MPB pH = 7,0	ASN	35	32	33
	CS	45	40	47
	BSS	43	40	45
MPB pH = 8,0	ASN	26	25	23
	CS	28	37	30
	BSS	35	30	22

The strain manifested strong antagonism against phytopathogenic fungi after culturing in all media. The spectrum of its antimicrobial action in different culture media was similar, but not identical. This suggests that the antibiotic substances produced by *B. methylotrophicus* 47 are probably synthesized in different amounts and may be similar in nature, but not the same. This strain is a promising biocontrol agent against phytopathogenic fungi.

The R_f - values of the antibiotic substances produced by *B. methylotrophicus* 47 were determined by thin layer chromatography. *B. methylotrophicus* 47 synthesized antibiotic substances, which determined its antifungal activity. In the thin layer chromatogram a spot with R_f value of 0.65, ninhydrin-positive was established so the synthesized antibiotic substances were peptides since the ninhydrin solution proved the presence of a protein ingredient (the stain was with the characteristic pink-violet color).

CONCLUSION

B. methylotrophicus 47 possesses high antifungal activity against *Aspergillus niger*, *Aspergillus awamori* and *Aspergillus flavus* after culturing in MPB or LBG-broth with different pH values under aerobic and anaerobic conditions. It was due to the synthesis of an antibiotic substance of a peptide nature with Rf value of 0.65.

REFERENCES

- [1] Leifert C., H. Li, S. Chidburee, S. Hampson, S. Workman, D. Sigeo, H. Epton, A. Harbour, Antibiotic production and biocontrol activity by *Bacillus subtilis* CL27 and *Bacillus pumilus* CL45, J Appl Bacteriol., 1995, 78(2), 97-108
- [2] Montesinos E., Antimicrobial Peptide and Plant Disease Control, FEMS Microbiology, Letters, 2007, 270, 1, 1-11
- [3] Niederwieser A., Thin-layer chromatography of amino acids and derivatives. Methods in Enzymology, 1972, 25, 60–99
- [4] Todorova S., Obtaining and characterization of a biopreparation with antibiotic action from *Bacillus subtilis* TS 01, Science & Technologies, Plant studies, 2014, 4(6), 217 – 221.
- [5] Yazgan A., G. Özcengiz, E. Özcengiz, K. Kılınç, M. A. Marahiel, N. G. Alaeddinoğlu, Bacilysin biosynthesis by a partially-purified enzyme fraction from *Bacillus subtilis*. Enzyme and Microbial Technology, 2001, 29(6–7), 400–406.

About the authors:

Rositsa Stefanova Denkova, PhD, Assistant professor at Department of Biochemistry and molecular biology, University of Food Technologies, Plovdiv, BG; e-mail: rositsa_denkova@mail.bg

Bogdan Georgiev Goranov, PhD, Department of Microbiology, University of Food Technologies, Plovdiv, BG; e-mail: dr.eng.bgoranov@gmail.com

Desislava Georgieva Teneva, PhD student at Department of Microbiology, University of Food Technologies, Plovdiv, BG; e-mail: desi_gerinska@yahoo.com

Zapryana Rangelova Denkova, DSc, Professor at Department of Microbiology, University of Food Technologies, Plovdiv, BG; e-mail: zdenkova@abv.bg

Hristo Marinov, Student, Department of Microbiology, University of Food Technologies, Plovdiv, BG; e-mail: hr_marinov@abv.bg

Georgi Atanasov Kostov, DSc, Associated professor at Department of Wine and Brewing, University of Food Technologies, Plovdiv, BG; e-mail: george_kostov2@abv.bg